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CALCIUM EFFLUX FROM CULTURED BOVINE ADRENAL CHROMAFFIN CELLS INDUCED BY BRADYKININ

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Abstract—The effect of bradykinin on Ca^{2+} efflux from cultured bovine adrenal chromaffin cells was examined. Bradykinin enhanced the efflux of $^{45}Ca^{2+}$ from the cells in a concentration dependent manner $(10^{-9}-10^{-6}\,\mathrm{M})$. This effect was inhibited by a specific bradykinin B_2 -receptor antagonist, but not by a B_1 -receptor antagonist. Nifedipine, Co^{2+} and Cd^{2+} did not inhibit the bradykinin-stimulated $^{45}Ca^{2+}$ efflux from the cells. 12-O-Tetradecanoyl phorbol 13-acetate, an activator of protein kinase C, also had no effect on the efflux of $^{45}Ca^{2+}$ from the cells. The increase in bradykinin-stimulated $^{45}Ca^{2+}$ efflux was reduced by removal of extracellular Na^+ . These results suggest that bradykinin stimulates Na^+/Ca^{2+} exchange in cultured bovine adrenal chromaffin cells.

The nonapeptide BK† influences several physiological processes including pain generation [1], blood pressure [2] and cardiovascular regulation [3]. It has also been suggested to be a central nervous system neurotransmitter and to play a role in the regulation of neuronal function [4, 5].

In neural cell lines, BK increases the level of cyclic GMP [6], stimulates production of inositol phosphates [7,8] and increases the cytosolic free Ca^{2+} concentration [9]. The increase in the intracellular free Ca^{2+} level, $[Ca^{2+}]_i$, was found to occur through agonist-stimulated influx of extracellular Ca^{2+} and hydrolysis of PIP₂ to yield IP₃ [10, 11].

Previously we reported that BK increases the intracellular levels of inositol phosphates, free Ca²⁺ and possibly diacylglycerol in pheochromocytoma PC-12 cells [12]. These effects increased the activities of protein kinases (calcium/calmodulin-dependent protein kinase and protein kinase C), resulting in stimulation of the pathway of catecholamine formation [12, 13]. Also in adrenal chromaffin cells. BK has been shown to stimulate the formation of inositol phosphates and to increase [Ca²⁺]_i. However, little is known of the mechanism of the fall in [Ca²⁺], elevation in adrenal chromaffin cells on BK stimulation. Hence the effect of BK on calcium efflux from cultured bovine adrenal chromaffin cells was studied. BK was found to enhance the efflux of ⁴⁵Ca²⁺ from these cells in culture and suggested that its effect may be mediated in part by acceleration of Na⁺/Ca²⁺ exchange.

MATERIALS AND METHODS

Cell preparation and culture. Bovine adrenal chromaffin cells were dispersed enzymatically as described previously [14]. Briefly, the medulla was sliced with a hand slicer, and the slices were digested in medium containing 0.1% collagenase, 0.01% soybean trypsin inhibitor, and 0.5% bovine serum albumin in BSS (135 mM NaCI, 5.6 mM KCI, 1.2 mM MgSO₄, 2.2 mM CaCI₂, 10 mM glucose and 20 mM HEPES/NaOH, pH 7.4). Cells were plated in 35-mm culture dishes at a density of 2×10^6 cells/ dish for measuring ⁴⁵Ca²⁺ efflux or on 22 × 22 mm cover glasses in 35-mm culture dishes at a density of 1×10^{6} cells/dish for measuring intracellular calcium, and maintained for 3 days as monolayer cultures in Eagle's basal medium supplemented with 5% heatinactivated fetal calf serum, 2 mM glutamine, penicillin (100 units/mL), streptomycin (100 $\mu g/mL$), gentamycin (40 $\mu g/mL$), fungizone (2.5) $\mu g/mL$) and 10 μM cytosine arabinoside.

⁴⁵Ca²⁺ efflux assay. Adrenal chromaffin cells were cultured in 35-mm dishes for 3 days. Then they were washed and incubated as described above in BSS containing ⁴⁵CaCI₂ (3 μ Ci/mL) for 1 hr at 37°. After incubation, the cells in each well were washed 15 times with 1 mL volumes of BSS at intervals for 30 sec to remove unincorporated ⁴⁵Ca²⁺. The cells were then incubated 15 times with 1 mL volumes of BSS for 30 sec periods to determine basal efflux levels. Then, they were incubated 15 times with 1 mL volumes of reaction mixture with or without test agents for 30 sec periods to determine agoniststimulated efflux levels. After agonist stimulation, the cells were solubilized in 1 mL 1% Triton X-100 to determine their residual ⁴⁵Ca²⁺. Samples were counted in 10 mL of liquid scintillation fluid for 2 min periods. The total radioactivity of 45Ca2+ in each well was determined as the sum of the radioactivity in each fraction and the residual radioactivity, and this value was used to calculate the fractional release of Ca²⁺ in each period.

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[†] Abbreviations: TPA, 12-O-tetradecanoyl phorbol 13-acetate; PIP₂, phosphatidyl inositol 4,5-bisphosphate; IP₃, inositol 1,4,5-triphosphate; BSS, balanced salt solution; BK, bradykinin.

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Measurement of intracellular calcium by fura-2. Intracellular Ca2+ level in single chromaffin cell was measured using the fluorescent Ca2+ indicator fura-2. The cells, which were cultured on cover glass, were incubated at 37° for 30 min with 1 mL BSS containing 2 μ M fura-2/acetoxy methyl ester. Then the cells on the cover glass were transferred to a small incubation bath (approx. 0.5 mL) on the platform of a microscope. The temperature was maintained at 37° and the incubation bath was perfused with BSS at a rate of 0.8 mL/min during the experimental period. Fluorescence was measured in single chromaffin cell on the cover glass using a fluorescence spectromicroscope (excitation, 340/380 nm, emission, 510 nm). The intracellular Ca²⁺ level was determined using the equation described previously [15].

Chemicals. ⁴⁵CaCl₂ was obtained from Amersham Corp. (Tokyo, Japan). BK, Des-Arg⁹-[Leu⁸]-BK and D-Arg-[Hyp³,Thi^{5,8},D-Phe⁷]-bradykinin were obtained from the Peptide Institute (Osaka, Japan). Nifedipine was obtained from Wako Pure Chemical Co. (Osaka, Japan). 12-O-Tetradecanoyl phorbol 13-acetate (TPA) and amiloride were obtained from Sigma Chemical Co. (St. Louis, MO, U.S.A.). Other chemicals used were commercial products of reagent grade.

RESULTS

Effects of bradykinin and BK antagonists on 45Ca²⁺ efflux

Figure 1 shows the effluxes of $^{45}\text{Ca}^{2+}$ from adrenal chromaffin cells in culture induced by various concentrations of BK. The stimulatory effect of BK on $^{45}\text{Ca}^{2+}$ efflux was dose-dependent at concentrations of 10^{-9} – 10^{-6} M BK. The efflux of $^{45}\text{Ca}^{2+}$ increased to a peak value within about 1 min after BK addition. The peak value with 10^{-6} M BK was $8.2 \pm 0.7\%$ (N = 6) of the total $^{45}\text{Ca}^{2+}$ in the cells. After the peak, efflux decreased rapidly in the next 5 min. Figure 2 shows the effects of the BK-receptor antagonists Des-Arg⁹-[Leu⁸] BK (B₁-receptor antagonist [16]) and D-Arg-[Hyp³, Thi^{5,8}, D-Phe⁷]-BK (B₂-receptor antagonist [17]) on the submaximal $^{45}\text{Ca}^{2+}$ efflux from the cells induced by 10^{-6} M BK. This efflux was inhibited 81% by 10^{-6} M D-Arg-[Hyp³, Thi^{5,8}, D-Phe⁷]-BK, but was not inhibited by Des-Arg⁹-[Leu⁸]-BK. This result suggests that the $^{45}\text{Ca}^{2+}$ efflux induced by bradykinin was mediated through the BK B₂-receptor.

Effects of various agents on 45Ca2+ efflux

To determine whether BK-stimulated ⁴⁵Ca²⁺ efflux is mediated by activation of Ca²⁺ channels, we examined whether it was inhibited by Ca²⁺ channel blockers. Nifedipine, an organic voltage-dependent Ca²⁺ channel blocker, had no effect on ⁴⁵Ca²⁺ efflux from the cells induced by BK (Fig. 3). The inorganic Ca²⁺ channel blockers, which inhibit voltage-dependent and receptor-operated Ca²⁺ channels, Co²⁺ and Cd²⁺ also did not inhibit BK stimulated ⁴⁵Ca²⁺ efflux (Fig. 3). Thus stimulation of ⁴⁵Ca²⁺ efflux by bradykinin is probably not due to increased Ca²⁺ flux through Ca²⁺ channels.

The stimulations of BK, histamine and muscarinic

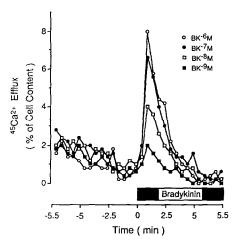


Fig. 1. Effects of different concentrations of BK on 45 Ca²⁺ efflux from cultured bovine adrenal chromaffin cells. Cells were preloaded with 45 Ca²⁺ as described in Materials and Methods, and then incubated with the indicated concentrations of bradykinin $(10^{-9}-10^{-6} \text{ M})$. Data are shown as percentages of cell contents, and are means for four to six separate experiments. Maximal SE was $\pm 10.6\%$.

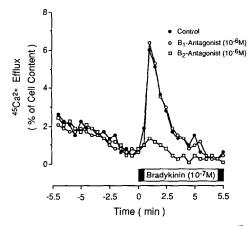


Fig. 2. Effects of BK antagonists on BK-induced ⁴⁵Ca²⁺ efflux from cultured bovine adrenal chromaffin cells. Cells were preloaded with ⁴⁵Ca²⁺ as described in Materials and Methods. The BK antagonist Des-Arg⁹-[Leu⁸]-BK (B₁-receptor antagonist; 10⁻⁶ M) or D-Arg-[Hyp³,Thi^{5,8},D-Phe⁷]-BK (B₂-receptor antagonist; 10⁻⁶ M), was added 150 sec before BK (10⁻⁷ M). Data are shown as percentages of cell contents, and are means for three to four separate experiments. Maximal SE was ±9.6%.

acetylcholine receptors are reported to induce breakdown of PIP_2 in cultured bovine adrenal chromaffin cells [18, 19]. The breakdown products of PIP_2 (IP_3 and diacylglycerol) increase the intracellular $[Ca^{2+}]_i$ level and stimulate protein kinase C [20]. The effects of bradykinin, histamine, acetylcholine and TPA (an activator of protein kinase C) on $^{45}Ca^{2+}$ efflux from the cells were

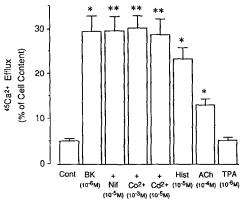


Fig. 3. Effects of various agents on $^{45}\text{Ca}^{2+}$ efflux from cultured bovine adrenal chromaffin cells. Cells were preloaded with $^{45}\text{Ca}^{2+}$ as described in Materials and Methods and then incubated in the presence or absence of BK (10 $^{-6}$ M), histamine (Hist; 10 $^{-5}$ M), acetylcholine (ACh; 10 $^{-4}$ M) or TPA (10 $^{-6}$ M). Nifedipine (Nif; 10 $^{-5}$ M), Co^{2+} (10 $^{-3}$ M) or Cd $^{+2}$ (10 $^{-5}$ M) was added 150 sec before BK (10 $^{-6}$ M). $^{45}\text{Ca}^{2+}\text{efflux}$ in 5 min after addition of agents was calculated. Data are means \pm SE for three to four separate experiments. * Significant difference from control (P < 0.01). ** No significant difference from BK stimulated $^{45}\text{Ca}^{2+}$ efflux.

examined (Fig. 3). Bradykinin $(10^{-6} \, \text{M})$ and histamine $(10^{-5} \, \text{M})$ increased the $^{45}\text{Ca}^{2+}$ efflux to about 460 and 350% the control level. Acetylcholine $(10^{-4} \, \text{M})$ increased $^{45}\text{Ca}^{2+}$ efflux from the cells slightly, but significantly, whereas TPA $(10^{-6} \, \text{M})$ had no effect on $^{45}\text{Ca}^{2+}$ efflux. These results suggest that bradykinin-stimulated $^{45}\text{Ca}^{2+}$ efflux from the cells is related to the formation of IP₃, but not to activation of protein kinase C.

Effects of BK, histamine and acetylcholine on intracellular free Ca²⁺ concentration

To determine whether the increased ⁴⁵Ca²⁺ efflux induced by BK is dependent on the elevation of intracellular [Ca²⁺]_i level, effects of BK, histamine and acetylcholine on intracellular free Ca²⁺ concentration were examined. As shown in Fig. 4, BK (10⁻⁶ M) and histamine (10⁻⁵ M) increased the intracellular [Ca²⁺]_i to approximately 430 and 360 nM. Acetylcholine (10⁻⁴ M) increased it to about 890 nM. However, acetylcholine-stimulated ⁴⁵Ca²⁺ efflux from the cells was less than BK-or histamine-stimulated ⁴⁵Ca²⁺ efflux (Fig. 3). Therefore, BK-stimulated ⁴⁵Ca²⁺ efflux from the cells may not be dependent on the elevation of intracellular [Ca²⁺]_i level in the cells.

Effects of extracellular sodium deprivation and amiloride on BK stimulated ⁴⁵Ca²⁺ efflux

To determine whether the increased ⁴⁵Ca²⁺ efflux induced by BK is Na⁺-dependent, we carried out a series of experiments in the absence of extracellular Na⁺. As shown in Fig. 5, complete replacement of Na⁺ by sucrose significantly blocked the enhanced ⁴⁵Ca²⁺ efflux from the cells induced by bradykinin.

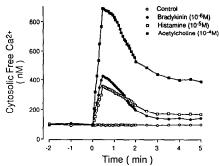


Fig. 4. Effects of BK, histamine and acetylcholine on intracellular free Ca²⁺ concentration in cultured bovine adrenal chromaffin cells. Cells were preloaded with 2 μ M fura-2/acetoxy methyl ester as described in Materials and Methods and then incubated in the presence or absence of BK (10⁻⁶ M), histamine (10⁻⁵ M) or acetylcholine (10⁻⁴ M). Data are means for four to five separate experiments. Maximal SE was $\pm 9.7\%$.

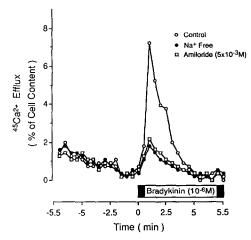


Fig. 5. Effects of Na⁺-free medium and amiloride on BK induced $^{45}\text{Ca}^{2+}$ efflux from cultured bovine adrenal chromaffin cells. Cells were preloaded with $^{45}\text{Ca}^{2+}$ as described in Materials and Methods. The medium was changed to Na⁺-free medium (with sucrose instead of all Na⁺) 150 sec before adding BK (10⁻⁶ M). In the case of the amiloride experiment, amiloride (5 × 10⁻³ M) was added 150 sec before BK (10⁻⁶ M) in normal BSS medium. Data are shown as percentages of cell contents, and are means for three to four separate experiments. Maximal SE was $\pm 11.8\%$.

Amiloride, an inhibitor of the Na⁺/Ca²⁺ exchanger [21], also significantly inhibited BK-stimulated ⁴⁵Ca²⁺ efflux from the cells. Therefore, the effect of BK in stimulating Ca²⁺ efflux across the plasma membrane may be mediated in part by a Na⁺/Ca²⁺ exchange mechanism.

DISCUSSION

In the present study we examined the mechanisms

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involved in stimulation of Ca2+ efflux from cultured bovine adrenal chromaffin cells by BK. BK is known to stimulate the formation of inositol phosphates in cultured bovine adrenal chromaffin cells [18, 19] and to increase $[Ca^{2+}]_i$ in a number of cell types [7, 8, 12, 22]. However, this increased $[Ca^{2+}]_i$ should be restored to a physiological level in response to further stimulus.

BK has been found to stimulate Ca2+ efflux from Swiss 3T3 fibroblasts [23], guinea pig tracheal cells [24] and human IMR-90 lung fibroblasts [25]. As shown in Fig. 1, it also increased Ca²⁺ efflux from cultured bovine adrenal chromaffin cells in a concentration-dependent manner. The concentration range of BK for this effect is similar to that for its stimulation of PIP₂ breakdown in the cells. We have reported that increases in inositol phosphates in pheochromocytoma PC-12 cells by BK is a B₂mediated response [12]. Here we report the pharmacological characterization of BK receptors in cultured bovine adrenal chromaffin cells determined by ⁴⁵Ca²⁺ efflux studies. The selective B₂-receptor antagonist D-Arg-[Hyp³,Thi^{5,8},D-Phe⁷]-BK inhibited the 45Ca2+ efflux from the cells induced by bradykinin, whereas the B₁-receptor antagonist Des-Arg⁹-[Leu⁸]-bradykinin [16] did not (Fig. 2). Thus this effect of BK may be mediated through the B2receptor.

The increased Ca2+ efflux induced by BK was not inhibited by nifedipine, Co²⁺ or Cd²⁺ (Fig. 3) suggesting that this efflux does not involve calcium channels in the cell membrane. In previous studies with pheochromocytoma PC-12 cells we found that BK had no effect on voltage-dependent calcium channels for Ca²⁺ uptake [12]. BK, therefore, does not seem to influence Ca²⁺ fluxes through calcium channels in adrenal chromaffin cells.

The increases in breakdown of PIP₂ in adrenal chromaffin cells induced by BK or histamine have been examined [18]. Diacylglycerol is produced concurrently with IP₃ on breakdown of PIP₂ by phospholipase C. Diacylglycerol is thought to activate protein kinase C by increasing the affinity of the enzyme for calcium [26-28]. Acetylcholine also induces production of IP₃ through the cholinergic muscarinic-receptor in adrenal chromaffin cells, though its efflux is less than these of BK and histamine [27]. In our study, BK and histamine treatments increased Ca2+ efflux from the cells approximately 5.6-and 4.5-fold, and acetylcholine increased it approximately 2.5-fold (Fig. 3). TPA, an activator of protein kinase C, had no influence on Ca²⁺ efflux from the cells (Fig. 3). Therefore, the production of IP3 induced by BK may regulate Ca²⁺ efflux from adrenal chromaffin cells.

The level of increase in $[Ca^{2+}]_i$ might influence Ca²⁺ efflux from the cells. However, treatments with 10⁻⁶ M BK, 10⁻⁵ M histamine and 10⁻⁴ M acetylcholine increased [Ca²⁺]_i in cultured bovine adrenal chromaffin cells to 430, 360 and 890 nM, respectively, as measured with fura-2 (Fig. 4). Thus the [Ca²⁺], after acetylcholine stimulation was higher than those after BK and histamine stimulations, whereas the increase in Ca2+ efflux from the cells after acetylcholine treatment was less than those after BK and histamine treatments (Figs 3 and 4).

Therefore, elevation of $[Ca^{2+}]_i$ did not apparently

increase Ca²⁺ efflux from adrenal chromaffin cells. Enhanced Ca²⁺ efflux from cultured bovine adrenal chromaffin cells following cell stimulation with a cholinergic agonist was found to depend on extracellular Na⁺ [29]. We observed enhanced ⁴⁵Ca²⁺ efflux from cultured bovine adrenal chromaffin cells during stimulation with BK and found here that this bradykinin-stimulated ⁴⁵Ca²⁺ efflux was inhibited in Na⁺-free medium and by amiloride treatment (Fig. 5). Thus BK potentiates Na⁺/Ca²⁺ exchange mechanisms in cultured bovine adrenal chromaffin cells. It is not clear if it is involved in some other BK-stimulated ⁴⁵Ca²⁺ efflux mechanisms because extracellular Na+ deprivation and amiloride could not completely abolish this efflux.

We conclude from this study that BK increases Ca2+ efflux from cultured bovine adrenal chromaffin cells, and that this efflux may be explained by stimulation of Na⁺/Ca²⁺ exchange mechanisms.

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